CLINICAL & FORENSIC

LC/MS/MS Analysis Using On-line Cartridges for the Removal of Phospholipids from Protein Precipitated Biological Fluid Samples

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Introduction

Phospholipids (PLs) are abundantly (at the mg/ mL level) present in biological fluids such as: blood, plasma, serum, and cerebrospinal fluids, among others. PLs are often co-extracted with a broad range of analytes of interest during sample preparation. The phospholipids present in a sample are notorious for producing various issues in LC/MS-based bioanalysis. PLs may cause ion suppression or, in rarer cases, ion enhancement, during MS detection. They also tend to build up on a reversed-phase (e.g., C18 and C8) column, fouling the chromatographic separation and ultimately shortening the column lifetime. Consequently, the accuracy, reproducibility, and sensitivity of LC/MS bioanalyses may be greatly compromised if the PLs are not removed.

HybridSPE® Phospholipid technology has been developed for selective and rapid depletion of phospholipids from biological samples prior to LC/MS analysis of small molecules. The technology utilizes the affinity of zirconia particles for selective binding and removal of phospholipids. The technology was introduced a few years ago in two product formats: 96-well filter plates for high throughput sample preparation and individual cartridges for low throughput applications. Here a new product format, an online cartridge, is described as an alternative option of phospholipid removal and sample preparation. The setup of the on-line cartridges with a LC/MS column is devised and the efficiency for phospholipid removal from protein precipitated plasma samples has been evaluated. Applicability of the system was demonstrated with three sets of compounds with different physiochemical properties.

Experimental

Materials

Supel[™] Genie HybridSPE[®] on-line cartridge (2 cm length x 4.0 mm i.d), Rat Plasma K2-EDTA (Lampire Cat. # 7306407); Protein precipitation solvent: Acetonitrile with 1% formic acid or methanol with 1% (w/v) ammonium formate.

Sample Processing Procedure: The rat plasma or rat plasma spiked with analytes was protein precipitated by vortex mixing the plasma samples with the precipitation solvent at a 1:3 ratio. The mixture was then centrifuged at 10,000 rpm x 3 min and the resulting supernatant was collected for LC/MS analysis.

Three sets of analytes Set-1: Basic analytes Risperidone Clomipramine Tamoxifen Set-2: Polar neutral analytes Digoxin Digitoxin Set-3: Non-polar neutral analytes 25-Hydroxyvitamin D₂ 3-epi-25-Hydroxyvitamin D₂ H₃C OH 3-epi-25-Hydroxyvitamin D₃ 25-Hydroxyvitamin D₃

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HybridSPE[®]-LC/MS setup

As shown in **Figure 2**, the setup consists of two pumps, one for HPLC separation, and the other for loading protein-crashed samples and washing the HybridSPE[®] on-line cartridges. A 50 μ L tee is employed for mixing of sample loading mobile phase and the HPLC separation mobile phases. The 2-position switching valve allows for washing the cartridges once the samples are loaded onto the HPLC column.



HybridSPE®-LC/MS conditions for Set-1 and -2 analytes

Instrument:	Shimadzu™ LCMS-8030 with 2DLC setup
HPLC column:	Ascentis [®] Express F5 10 cm x 2.1 mm (53569-U)
Mobile phase:	(A) Water/10 mM ammonium formate;(B) methanol
Gradient:	0% B% for 4 min, to 75% B in 0.5 min, held for 8 min
Flow:	0.3 mL/min
Column temperature:	45 °C
Sample loading flow:	0.1 mL/min
Sample loading solvent:	methanol with 10 mM ammonium formate
Injection Vol:	1 μL
Detection:	MS, ESI(+), MRM mode

HybridSPE®-LC/MS conditions for Set-3 analytes

Instrument:	Shimadzu ^{m} LCMS-8030 with 2DLC setup
HPLC column:	Ascentis [®] Express C18 5 cm x 2.1 mm (53822-U)
Mobile phase:	(A) Water; (B) 90% Acetonitrile, each with 10 mM ammonium formate
Gradient:	0% B% for 4 min, to 80% B in 2 min, held for 1.5 min
Flow:	0.3 mL/min
Column temperature:	35 °C
Sample loading flow:	0.1 mL/min
Sample loading solvent:	80% acetonitrile with 50 mM ammonium formate
Injection Vol:	1 μL
Detection:	MS, ESI(+), MRM mode

HybridSPE®-LC/MS conditions for phospholipid detection:

Instrument:	Shimadzu [™] LCMS-8030 with 2DLC setup
HPLC column:	Ascentis [®] Express OH5 5 cm x 2.1 mm (53749-U)
Mobile phase:	(A) Water; (B) 90% Acetonitrile, each with 10 mM ammonium formate
Isocratic:	85% B
Flow:	0.3 mL/min
Column temperature:	35 °C
Sample loading flow:	0.2 mL/min
Sample loading solvent:	90% acetonitrile with 10 mM ammonium formate
Injection:	1 μL
Detection:	MS, ESI(+), MRM mode
Phospholipid ions:	precursor ions 496, 520, 522, 524, 758, 782, 786, and 810, product ions are all 184

Results and Discussion

The SupelTM Genie HybridSPE[®] on-line cartridges are designed for removal of phospholipids from more than 100 injections of 1 µL of protein precipitated plasma samples. **Figure 3**(A) shows the phospholipids in the rat plasma, if not removed, give rise to two broad peaks of high intensity. The two peaks correspond to the phospholipids containing one and two fatty acyl chain(s), respectively. When the HybridSPE[®] on-line cartridge is set up with the LC/MS (**Figure 2**), no phospholipid peaks were detected, even at the 120th injection of the same rat plasma sample (see **Figure 3**(B)). The results demonstrate the HybridSPE[®] cartridges are capable of elimination of the phospholipids from 120 consecutive injections of 1 µL of protein precipitated plasma samples.

Figure 3. (A) Phospholipids in plasma sample without a HybridSPE[®] cartridge; (B) #120th injection of the same plasma sample with a HybridSPE[®] on-line cartridge set up with LC/MS (see Figure 2 for the setup).



The applicability of the HybridSPE® on-line cartridges has been demonstrated with three sets of analytes including basic and neutral classes. As can be seen from Figure 4-6, narrow and symmetric peaks were observed for all of the tested analytes with a peak

width at half height <6 s and tailing factors 0.9-1.3, respectively. Table 1 summarizes the recoveries and reproducibilities of the target compounds using the on-line SPE method. For all of the tested analytes, recoveries of 94-102% were obtained and reproducibilities of 1-5% were achieved.



Peak	Analyte	Peak width at 50% height (s)	Taliling factor
1	Risperidone	3.54	1.2
2	clomipramine	3.78	1.3
3	Tamoxifen	3.36	1.2

- All peaks are narrow: <4s peak width at
- Both peaks are symmetric, with tailing
- Baseline is low and clean: no interference





Peak	Analyte	Peak width at 50% height (s)	Taliling factor
1	Digoxin	2.52	1.3
2	Digitoxin	2.70	1.2

- Both peaks are narrow: <3s peak width at half height
- Both peaks are symmetric, with tailing factors of 1.2-1.3
- Baseline is low and clean: no interference peaks



Figure 6. Representative LC/MS chromatogram of non-polar neutral (Set-3) analytes

Peak	Analyte	Peak width at 50% height (s)	Taliling factor
1	25-OH D3	4.74	0.9
2	3- <i>epi</i> -25-OH D3	4.80	0.9
3	25-OH D2	4.62	1.0
4	3- <i>epi</i> -25-OH D2	5.10	0.9

- All peaks are narrow: <6s peak width at half height
- Both peaks are symmetric, with tailing factors of 0.9-1.0
- Baseline is low and clean: no interference peaks

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Table 1. Analyte's Recovery and Reproducibility

Analyte	Retention time (mm	MRM Quantifier	Avg. Recovery*	%RSD, n=20
Digoxin	5.4	798.5/651.5	96%	4.9
Digitoxin	5.9	782.5/243.2	97%	2.2
Risperidone	5.4	411.3/191.3	102%	1.5
clomipramine	6.0	315.2/86.1	94%	1.1
Tamoxifen	6.6	372.3/72.2	98%	1.4
25-OH D3	8.2	401.4/383.3	102%	1.2
3- <i>epi</i> -25-OH D3	8.4	401.4/383.3	102%	2.5
25-OH D2	8.4	413.4/395.4	100%	2.0
3- <i>epi</i> -25-OH D2	8.6	413.4/395.4	99%	4.2

* The recovery was calculated by comparison of the peak area of the spiked analytes in plasma to those of the neat analytes at the same concentration.

Summary

An on-line cartridge for phospholipid removal with LC/MS analysis has been successfully developed. The performance testing demonstrates the on-line cartridges are capable of removing >95% of phospholipids from 1 μ L of plasma samples even after 120 consecutive injections. Three applications have been established using on-line HybridSPE[®] with LC/MS detection. Recovery of the analytes is 94%-102%, with a reproducibility of 1%-5%. For all of the tested analytes, narrow and symmetric peaks were observed, peak width at half height <6 s and tailing factors 0.9-1.3, respectively.

Related Products

Description	Cat. No
Sample Preparation	
Supel [™] Genie RP-Amide On-line Starter Kit	55516-U
Supel™ Genie RP-Amide On-line SPE Cartridge, pk. of 2	55519-U
Supel™ Genie RP-Amide On-line SPE Cartridge, pk. of 6	55522-U
Supel™ Genie C8 On-line Starter Kit	55274-U
Supel [™] Genie C8 On-line SPE Cartridge, pk. of 2	55512-U
Supel [™] Genie C8 On-line SPE Cartridge, pk. of 6	55515-U
Solvents	
Water for chromatography (LC-MS Grade) LiChrosolv $\ensuremath{^{\$}}$	1.15333
Acetonitrile hypergrade for LC-MS LiChrosolv [®]	1.00029

Featured Products

Description	Cat. No.
Sample Preparation	
Supel [™] Genie HybridSPE [®] On-line Starter Kit	55324-U
Supel [™] Genie HybridSPE [®] On-line SPE Cartridge, pk. of 2	55326-U
Supel [™] Genie HybridSPE [®] On-line SPE Cartridge, pk. of 6	55327-U
LC	
Ascentis® Express C18 5 cm x 2.1 mm, 2.7 µm	53822-U
Ascentis® Express F5 10 cm x 2.1 mm, 2.7 µm	53569-U
Ascentis [®] Express OH5 5 cm x 2.1 mm, 2.7 µm	53749-U

Did you know...

 \ldots that the HybridSPE $^{\otimes}$ material is also available in 96-well plates for protein & phospholipid removal in one off-line step?

Read more under SigmaAldrich.com/hybridspe



SPME LC (BioSPME) Tips

Microsampling and Sample Preparation in one step



SigmaAldrich.com/biospme

